Aetiology of Acute Febrile Episodes in Children Attending Korogwe District Hospital in North-Eastern Tanzania

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Abstract

Introduction: Although the burden of malaria in many parts of Tanzania has declined, the proportion of children with fever has not changed. This situation underscores the need to explore the possible causes of febrile episodes in patients presenting with symptoms at the Korogwe District Hospital (KDH).

Methods: A hospital based cross-sectional study was conducted at KDH, north-eastern Tanzania. Patients aged 2 to 59 months presenting at the outpatient department with an acute medical condition and fever (measured axillary temperature \( \geq 37.5^\circ C \)) were enrolled. Blood samples were examined for malaria parasites, human immunodeficiency virus (HIV) and bacterial infections. A urine culture was performed in selected cases to test for bacterial infection and a chest radiograph was requested if pneumonia was suspected. Diagnosis was based on both clinical and laboratory investigations.

Results: A total of 867 patients with a median age of 15.1 months (Interquartile range 8.6–29.9) were enrolled from January 2013 to October 2013. Respiratory tract infections were the leading clinical diagnosis with 406/867 (46.8%) of patients diagnosed with upper respiratory tract infection and 130/867 (15.0%) with pneumonia. Gastroenteritis was diagnosed in 184/867 (21.2%) of patients. Malaria infection was confirmed in 72/867 (8.3%) of patients. Bacterial infection in blood and urine accounted for 26/808 (3.2%) infections in the former, and 66/373 (17.7%) infections in the latter. HIV infection was confirmed in 10/824 (1.2%) of patients. Respiratory tract infections and gastroenteritis were frequent in patients under 36 months of age (87.3% and 91.3% respectively). Co-infections were seen in 221/867 (25.5%) of patients. The cause of fever was not identified in 65/867 (7.5%) of these patients.

Conclusions: The different proportions of infections found among febrile children reflect the causes of fever in the study area. These findings indicate the need to optimise patient management by developing malaria and non-malaria febrile illnesses management protocols.


Introduction

More than half of the children presenting at health facilities in Africa are estimated not to have a malaria infection [1,2]. The lack of knowledge on the prevalence of the causative agents of other febrile illnesses, the limited access to affordable diagnostic tests as well as the fear of overlooking a life threatening malaria infection, result in most febrile cases being treated and managed as malaria [3,4].

Fever commonly accompanies a wide range of childhood illnesses and if the patient is not properly managed, this can have potentially serious outcomes including death [5]. In malaria endemic countries, febrile episodes are commonly associated with malaria infection [6,7]. As current epidemiological data indicates a decline in malaria infections in many parts of sub Saharan Africa, including Tanzania, other causative agents of febrile illnesses should be investigated [8,9].

Acute febrile episodes are caused by various pathogenic organisms (such as viruses and bacteria), and infections with these agents result in patients presenting with clinical symptoms which resemble those of a malaria infection [10]. An accurate clinical diagnosis without laboratory confirmation can be difficult to make and misleading [11,12]. Despite efforts by the World Health Organisation (WHO) to improve the diagnosis of malaria through its updated integrated management of childhood illness (IMCI)
guidelines, confirmatory diagnosis of malaria is still not adequately carried out. In routine practice, clinicians have continued prescribing antimalarial drugs to malaria negative patients [13,14]. As IMCI approach is apparently not specific on the diagnosis of non-malarial febrile illnesses [15]. This results in the inappropriate use of antimalarial drugs, possible development of antimicrobial drug resistance and also elevates treatment costs due to unnecessary prescriptions. Incorrect prescriptions may leave patients untreated, thereby unnecessarily increasing morbidity and mortality in these resource poor settings [16].

There is limited data on the epidemiological causes of febrile illnesses (other than malaria) among outpatient children, as well as a lack of reliable surveillance data in Tanzania. A recent study conducted in Tanzania found that malarial infections were no longer a major cause of fever among outpatient children [17]. Disease-specific studies among febrile patients have been conducted in few settings across Sub Saharan Africa. These studies focused mainly on malaria, bacteraemia and human immunodeficiency virus (HIV) infection among inpatients [18–20].

Information on the prevalence of local infections, that is based on diagnosis using both clinical presentation and laboratory confirmatory tests is critical for correct management of both malarial and non-malarial febrile illnesses. The main objective of this study was to describe the common causes of fever in children presenting at an outpatient department at the Korogwe District Hospital, north-eastern Tanzania.

Methods

Study area

The study was conducted at Korogwe District Hospital (KDH) in Korogwe District, Tanga Region, north-eastern part of Tanzania. The Korogwe District has a population of approximately 73,275 children under the age of five years according to the Tanzanian population and housing census of 2012 [21]. The District has a population growth rate of 2.7% per annum [21]. The environment is characterised by daily temperatures varying from 18°C to 20°C during the rainy season and 26°C to 30°C during the dry season. The annual rainfall ranges from 700–1000 mm with long rainy seasons extending from March to May. The majority of the inhabitants reside in rural settings, practicing subsistence farming and informal trade. The hospital receives 6000 (2010 estimates) children under the age of five years as outpatients annually. The most common clinical diagnoses among children under the age of five years have been malaria, pneumonia, gastroenteritis, septicaemia, anaemia and diarrhoea (Korogwe District Medical Officer, personal communication).

The prevalence of Plasmodium falciparum malaria parasitaemia from the community in lowland villages has decreased from 78% in 2003 to 13% in 2008, whereas in the highland villages, the decrease was from 25% to 3% during the same time period [22]. The prevalence of HIV infection among women attending the antenatal clinic at KDH was 2.5% for 2010 [23]. Pneumococcal and rotavirus vaccines were introduced into Tanzania in January 2013 as part of the Expanded Program on Immunization. Vaccine coverage for other vaccines in the Expanded Program on Immunization (EPI) has been above 80%. These vaccines include; Bacille Calmette-Guérin, pentavalent vaccine (diphtheria–tetanus–pertussis, hepatitis B and haemophilus influenza type B), poliovirus and measles vaccines.

Study population

Sick children aged between 2 and 59 months presenting at KDH outpatient department were assessed for study eligibility from January 2013 to October 2013. Enrolment took place Mondays to Fridays every week. The inclusion criteria were: children aged 2 to 59 months of age presenting at KDH with an acute medical condition and a history of fever in the last 48 hours or measured axillary temperature of ≥37.5°C. The visit should be their first consultation for the present problem. Patients younger than 2 months of age were excluded in the study. Patients were excluded if they had planned admissions (e.g. elective surgery), trauma/injury, if they required an emergency intervention or if they had taken antimalarial drugs and/or antibiotics within the last seven days.

Ethics statement

The study obtained ethical clearance with reference number NIMR/HQ/R.8a/Vol.IX/1373 from the Tanzanian Medical Research Coordinating Committee. A written informed consent form was issued and signed by the parent/legal guardian of every child enrolled in the study.

Study procedure

A detailed medical history and a thorough clinical examination were performed on each patient and this information was recorded on a standardised Case Report Form (CRF). This information included: demographic information, clinical history and physical examination data (including weight, axillary temperature and respiratory rate). A chest radiograph was requested if pneumonia was suspected. A clinical diagnosis was made based on the presenting signs and symptoms according to the IMCI guidelines [24]. This ensured that patients could be appropriately managed according to national standard practice while laboratory tests were being processed. Patients who had positive culture results were contacted and the necessary required treatment provided. The Final diagnosis was based on both the clinical presentation and the laboratory findings for malaria microscopy, bacteriology, radiology and HIV testing.

Laboratory investigations

A maximum of 5 mL venous blood for laboratory investigations was drawn from every patient once inclusion criteria had been met. Each blood draw was conducted aseptically in order to avoid contamination. The venipuncture site was disinfected with 70% isopropyl alcohol and allowed to dry prior to blood drawing.

Diagnosis of Plasmodium species and Borrelia parasitic infection

Thick and thin blood smears were prepared (in duplicate) from the blood collected in the ethylene diamine tetra acetic acid (EDTA) tubes. The thin film was fixed with methanol and blood slides were stained with a 5% Giemsa solution for identification and quantitation of asexual Plasmodium falciparum and other Plasmodium species. The blood slides were read by two expert microscopists and in case of discrepancy, a third reading was performed. Asexual parasites were counted against 200 (500 if parasite count was <10) white blood cells. A blood slide was considered negative for Plasmodium species if no parasites were seen in at least 100 oil-immersion high power fields on the thick film. During examination of the slides for Plasmodium species, Borrelia species were also screened. Borrelia species are seen as spirochaetes under microscopy.

Differential complete blood count

Blood collected in anticoagulated EDTA tubes was analysed for differential complete blood count using automated MS4s haematology analyser (Melet Schloesing Diamond Diagnostics, USA).
Aerobic blood culturing

Blood collected in commercially available BD BACTEC PEDS PLUS culture bottles were incubated in an automated blood culture system BACTEC 9050 (Becton Dickinson, Sparks, Maryland, USA). A single blood culture bottle was used per patient. The minimal volume inoculated was 2 mL (3 mL was an ideal volume) and blood culture bottles were incubated for a maximum of five days (unless they flag positive). The BACTEC 9050 detects positive cultures based on CO₂ production. Blood culture bottles which flagged positive were cultured on standard media with the use of routine microbiological techniques. Analytical Profile Index (API) biochemical test kit (BioMérieux, France) and/or serological tests were used to confirm suspected pathogens.

Definition of positive and negative culture

A blood culture was considered positive when: a definite pathogen was isolated (e.g. Streptococcus pneumoniae, Streptococcus agalactiae, Streptococcus pyogenes, Haemophilus influenzae, Salmonella species), a bacteria that could be either a pathogen or a contaminant was isolated within 48 hours of blood culture incubation (e.g. Escherichia coli, Klebsiella pneumoniae, Staphylococcus aureus, Enterococcus faecalis group D). Blood cultures were considered to be negative if there were no bacteria isolated after five days incubation.

Definition of contamination

A blood culture was considered to be contaminated if one or more of the following organisms were identified: coagulase-negative Staphylococcus species, Corynebacterium species, alphahemolytic streptococci, Micrococcus species, Bacillus species and Propionibacterium species. However, a definitive diagnosis of a contaminant was decided by a clinician based on the clinical presentation of the patient due to the fact that these organisms could be the cause of an opportunistic infection.

Urine culturing

Urine was collected from patients presenting with fever, dysuria and/or no obvious cause of fever on clinical examination. A midstream urine was collected in a sterile container. The perineum was swabbed with chlorhexidine and the sample was collected using a sterile urine collector for the younger patients who were not able to follow instructions. Urinalysis by dipstick (URiScan, YD Diagnostics, Korea) was used for presumptive diagnosis of urinary tract infections for early initiation of therapy while urine cultures were being processed. The urine was cultured by inoculation onto cysteine lactose electrolyte deficient agar, MacConkey agar and sheep blood agar. An API biochemical test kit (BioMérieux, France) was used to confirm suspected pathogens. A definitive diagnosis of urinary tract infection was based on the isolation of a bacteria pathogen from the positive culture containing \(10^5\) colony-forming units (CFU)/ml [25].

Antimicrobial susceptibility testing

The Kirby-Bauer disc diffusion method was used for in-vitro antimicrobial susceptibility testing for all pathogenic bacterial isolates from blood and urine cultures. The testing was performed on the following antimicrobial agents; amoxicillin, chloramphenicol, ceftriaxone, ciprofloxacin, co-trimoxazole, gentamicin and penicillin. Reading and interpretation of zone sizes was performed using criteria stipulated by the Clinical and Laboratory Standards Institute (CLSI).

Human immunodeficiency virus (HIV) testing

HIV antibody testing was performed after patients were informed and counselled as per the National Health Guidelines. Blood drawn from patients was tested for the presence of HIV-1 and HIV-2 antibodies according to the National HIV rapid testing algorithm using the Determine HIV-1/2 Assay Kit (Abbott Laboratories, Abbott Park, IL, USA) [26]. If this was positive, confirmation was carried out using the Uni-Gold HIV (Trinity Biotech, Bray, Co Wicklow, Ireland). For children under the age of 18 months, HIV testing was performed using HIV-1 RNA PCR.

Case definitions

Fever was defined as history of abnormally high body temperature as reported by the parent/guardian and/or measured axillary temperature \(\geq 37.5°C\) on presentation. Malaria infection was defined as fever with the presence of asexual Plasmodium falciparum parasites in a blood smear confirmed by microscopy. Bacteremia was defined as fever with isolation of pathogenic bacteria from blood culture. Anaemia was defined as haemoglobin concentration below 9.3 g/dL. Clinical diagnoses of respiratory infections and gastroenteritis were defined according to the IMCI guidelines [24].

Data management and statistical analysis

Data management involved double entry and validation using Microsoft Access 2007. Data analysis was done by STATA version 11.2 (Stata Corp LP, College Station, Texas, USA). Variables were summarized as frequencies and proportions and medians and inter-quartile ranges, as appropriate. Data was compared using the chi-square (\(X^2\)) test when comparing proportions. Odds ratios (OR) and 95% confidence intervals (95% CI) were calculated as appropriate. The value of \(p<0.05\) was considered statistically significant.

Results

Demographic characteristics of the study population

A total of 1380 patients attending the outpatient department at KDH from January 2013 to October 2013 were screened. Of these, 867/1380 (62.8%) were febrile children who were eligible for enrolment (Figure 1). The median age was 15.1 months (Interquartile range (IQR): 8.6–29.9) and girls comprised of 439/867 (52.9%) patients as shown on Table 1. Median axillary body temperature was 30.1°C (IQR: 29.3–30.7). The common presenting clinical symptoms other than fever were cough 411/867 (47.4%) and diarrhoea 174/867 (20.1%).

Malaria infection

Malaria infection due to Plasmodium falciparum was confirmed in 72/867 (8.3%, 95%CI: 6.5–10.1) of the patients identified by thick and thin blood smears. Anaemia was found in 226/841 (26.9%) patients, of whom 36/226 (15.9%) had malaria which increased the risk of anaemia (OR: 3.6 [95%CI: 2.1–6.1] \(p<0.01\)). No other Plasmodia species or Borrelia species were found. Overlap of clinical symptoms was observed among patients with malaria infection; 14/72 (19.4%) and 4/72 (5.6%) patients had clinical symptoms of respiratory tract infections and gastroenteritis respectively.

Bacteremia

Blood cultures were collected and analysed from 808 patients, of which 26/808 (3.2%, 95%CI: 1.9–4.4) were positive for pathogenic bacterial growth. Salmonella typhi was the predominant
bacteria isolated in 17/26 (65.4%) of cases followed by *Streptococcus pneumoniae* 4/26 (15.4%) as shown in Table 1.

**Urinary tract infection**

A urinary tract infection was confirmed in 66/373 (17.7%, 95%CI, 14.0–22.0) of patients and there was no significant association with gender (36 boys, 30 girls, $X^2 = 0.074, P = 0.786$). The most commonly isolated bacteria were *Escherichia coli* 37/66 (56.1%) followed by *Klebsiella pneumoniae* 7/66 (10.6%), *Staphylococcus aureus* 6/66 (9.1%) and *Proteus mirabilis* 5/66 (7.6%).

**HIV infection**

HIV testing was performed in 824/867 (95.0%) patients and among them, 10/824 (1.2%, 95%CI, 0.6–2.2) were confirmed positive for infection with HIV. None of the HIV infected patients had either a malarial or a bacterial infection.

**Clinical diagnoses**

**Respiratory tract infections.** Upper respiratory tract infections were the most common clinical diagnoses presenting in 406/867 (46.8%, 95%CI, 43.5–50.2) of patients. Pneumonia was clinically diagnosed in 130/867 (15.0%, 95%CI, 12.7–17.5) of patients. Pneumonia confirmed by chest radiograph was accounted in 54/130 (41.5%) of these cases and five patients had diagnosis of severe pneumonia. *Salmonella typhi* ($n = 2$) and *Escherichia coli* ($n = 1$) were isolated from single blood cultures of patients with upper respiratory tract infections. *Streptococcus pneumoniae* ($n = 1$) and *Staphylococcus aureus* ($n = 1$) were isolated from single blood cultures of patients with pneumonia diagnosis. One patient was suspected to have pulmonary tuberculosis from clinical symptoms and chest radiograph results.

**Gastroenteritis.** Gastroenteritis was diagnosed in 184/867 (21.2%, 95%CI, 18.5–24.1) of patients. Dysentery was diagnosed in 6 patients. *Salmonella typhi* ($n = 6$) and non-typhi salmonella ($n = 1$) were isolated from the blood cultures of patients with gastroenteritis.

**Other infections.** Other infections are summarised on Table 1. The cause of fever could not be identified in 63/867 (7.5%, 95%CI, 5.8–9.5) of the patients.

**Distribution of diagnoses according to age.** The majority of the patients under the age of 36 months presented with respiratory tract infections 468/536 (87.3%). The second most common diagnosis was gastroenteritis 168/184 (91.3%) and thirdly, urinary tract infections 57/66 (86.4%). Malaria infection 8/72 (11.1%) was less common in patients under the age of 12 months (Figure 2).

**Diagnosis of hospitalised patients**

Following enrolment, 141/867 (16.3%) patients were hospitalised, of whom 2/141 (1.4%) died from severe pneumonia and upper respiratory tract infection. Malaria infection was a leading cause of admission with 53/141 (37.6%) of patients, followed by pneumonia 35/141 (24.6%), gastroenteritis 27/141 (19.1%) and other infections as indicated in Figure 3.

**Co-infections**

Co-infections were common and were observed in 221/867 (25.5%, 95%CI, 22.6–28.5) of the patients. Urinary tract
infections were present in 33/436 (7.6%) patients with respiratory tract infections and in 19/184 (10.3%) patients with gastroenteritis (Figure 4). Five patients had multiple diagnoses of urinary tract infection, respiratory tract infection and gastroenteritis. Among patients with malaria, 8/72 (11.1%) patients had urinary tract infections. Sixty patients had both respiratory tract infections as well as gastroenteritis.

Table 1. Demographic characteristics, clinical diagnosis and laboratory findings of patients enrolled at KDH.

<table>
<thead>
<tr>
<th>Patient characteristics</th>
<th>n, n/N</th>
<th>Proportions (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Gender</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Girls</td>
<td>408</td>
<td>47.1</td>
</tr>
<tr>
<td><strong>Age</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2–11 months</td>
<td>356</td>
<td>41.1</td>
</tr>
<tr>
<td>12–35 months</td>
<td>370</td>
<td>42.7</td>
</tr>
<tr>
<td>36–59 months</td>
<td>141</td>
<td>16.3</td>
</tr>
<tr>
<td><strong>Fever</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Duration of fever in days, median (IQR)</td>
<td>2 (1–2)</td>
<td>-</td>
</tr>
<tr>
<td>Axillary temperature ≥37.5 °C</td>
<td>627</td>
<td>72.3</td>
</tr>
<tr>
<td><strong>Common symptoms</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cough</td>
<td>411</td>
<td>47.4</td>
</tr>
<tr>
<td>Diarrhoea</td>
<td>174</td>
<td>20.1</td>
</tr>
<tr>
<td>Difficulty breathing</td>
<td>97</td>
<td>11.2</td>
</tr>
<tr>
<td><strong>Clinical diagnosis</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Respiratory infections</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Upper respiratory tract infection</td>
<td>406/867</td>
<td>46.8</td>
</tr>
<tr>
<td>Pneumonia</td>
<td>130/867</td>
<td>15</td>
</tr>
<tr>
<td>Pulmonary Tuberculosis</td>
<td>1/867</td>
<td>0.1</td>
</tr>
<tr>
<td>Asthma</td>
<td>19/867</td>
<td>2.2</td>
</tr>
<tr>
<td>Gastroenteritis</td>
<td>184/867</td>
<td>21.2</td>
</tr>
<tr>
<td>Dysentery</td>
<td>6/867</td>
<td>0.7</td>
</tr>
<tr>
<td>Otitis media</td>
<td>5/867</td>
<td>0.6</td>
</tr>
<tr>
<td>Fungal infections</td>
<td>6/867</td>
<td>0.7</td>
</tr>
<tr>
<td>Skin infections/soft tissue infections</td>
<td>33/867</td>
<td>3.8</td>
</tr>
<tr>
<td>Other infections</td>
<td>44/867</td>
<td>5.1</td>
</tr>
<tr>
<td><strong>Laboratory findings</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Plasmodium falciparum</td>
<td>72/867</td>
<td>8.3</td>
</tr>
<tr>
<td>Geometric mean parasitemia (range)</td>
<td>37635 (159–1656400)</td>
<td>-</td>
</tr>
<tr>
<td>Human immunodeficiency virus</td>
<td>10/824</td>
<td>1.2</td>
</tr>
<tr>
<td>Anaemia</td>
<td>226/841</td>
<td>26.9</td>
</tr>
<tr>
<td><strong>Blood culture positive</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Non-typhi salmonella</td>
<td>2/26</td>
<td>7.7</td>
</tr>
<tr>
<td>Salmonella typhi</td>
<td>17/26</td>
<td>65.4</td>
</tr>
<tr>
<td>Streptococcus pneumoniae</td>
<td>4/26</td>
<td>15.4</td>
</tr>
<tr>
<td>Other¹</td>
<td>3/26</td>
<td>11.5</td>
</tr>
<tr>
<td><strong>Urine culture</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Escherichia coli</td>
<td>37/66</td>
<td>56.1</td>
</tr>
<tr>
<td>Klebsiella pneumoniae</td>
<td>7/66</td>
<td>10.6</td>
</tr>
<tr>
<td>Proteus mirabilis</td>
<td>5/66</td>
<td>7.6</td>
</tr>
<tr>
<td>Other²</td>
<td>17/66</td>
<td>25.7</td>
</tr>
<tr>
<td><strong>Pyrexia of Unknown Origin</strong></td>
<td>65/867</td>
<td>7.5</td>
</tr>
</tbody>
</table>

¹Escherichia coli (1), Enterobacter cloacae (1), Staphylococcus aureus (1).
²Acinetobacter baumannii (1), Citrobacter koseri (1), Pseudomonas aeruginosa (2), Staphylococcus aureus (6) Staphylococcus saprophyticus (3), Streptococcus faecalis (1), Streptococcus mitis (1), Streptococcus viridans (2).

doi:10.1371/journal.pone.0104197.t001
Figure 2. Distribution of common diagnosis according to age group.
doi:10.1371/journal.pone.0104197.g002

Figure 3. Distribution of illnesses among hospitalised patients at KDH (n = 141).
doi:10.1371/journal.pone.0104197.g003
Antimicrobial susceptibility pattern

Gram negative bacterial isolates demonstrated good susceptibility to ciprofloxacin (80%-100%), moderate susceptibility to gentamicin (50%-72%) and poor susceptibility to amoxicillin, chloramphenicol and co-trimoxazole (3%-14%). *Escherichia coli* and *Klebsiella pneumoniae* demonstrated poor susceptibility to ceftriaxone (35.1% and 14.3% respectively). *Klebsiella pneumoniae* was the only Gram negative bacteria with good susceptibility to chloramphenicol (71.4%). The Gram positive bacterial isolates were observed to have good susceptibility to ceftriaxone and chloramphenicol (75%-100%) and poor susceptibility to co-trimoxazole and penicillin (0%-50%).

Discussion

This study has been able to contribute information on the prevalence of the possible causes of common febrile episodes among children less than five years of age, attending an outpatient department at KDH in north-eastern Tanzania. Over three quarters of children presenting with fever were under the age of 36 months and the majority had respiratory tract infections and gastroenteritis as the leading clinical diagnosis. Younger children under the age of 36 months tend to be more vulnerable to infections due to their immature immunity, different modes of exposure to pathogenic organisms and possibly due to a low rate of immunisation [27,28].

*Plasmodium falciparum* malaria, invasive community-acquired bacteria and HIV infections were uncommon among febrile children in the study with a prevalence of 8.3%, 3.2% and 1.2%, respectively. These findings are consistent with other studies on the declining malaria and community-acquired bacterial infections in the region [29,30]. The low malaria prevalence rate might partly be due to current interventions which include use of insecticide treated nets, the wide availability of effective antimalarial drugs, residual spraying and change of malaria vector behaviour [31,32]. This underscores the importance of the rational use of antibiotic and antimalarial drugs in evidence based medicine among less severely ill patients in order to counteract the problem of drug resistance.

This study found an increasing prevalence of *Salmonella typhi* among young children compared with that of non-typhi salmonella. These findings are in keeping with those of Biggs and colleagues, who found similar changing pattern of bacterial infections where *Salmonella typhi* compared with non-typhi salmonella was more common in lower malaria prevalent areas [20]. The observed prevalence of *Salmonella typhi* indicates the presence of typhoid fever among these young rural patients. The following critical measures can be implemented in preventing typhoid fever in this resource poor area through improving the quality of life. This can be done by ensuring good sanitation, safe water supply and public health education on personal as well as food hygiene.

Amoxicillin, chloramphenicol and co-trimoxazole are the most widely available antimicrobial agents in the community. These were shown to demonstrate poor susceptibility to the leading Gram negative bacterial isolates from both blood and urine cultures. Ciprofloxacin was effective but not recommended for use in children under the age of five years [33,34]. The findings support previously published work and emphasize the increasing problem of antimicrobial drug resistance [35,36]. Specific identification of bacteria causing infections is important for understanding and monitoring of antimicrobial drug resistance patterns. This will ensure patients especially children are provided with appropriate and effective treatment.

The prevalence of urinary tract infections in this study was higher among children under 36 months of age and these patients were often co-infected. The lack of access to clean water and good sanitation in this rural community might be the reason for the higher prevalence of infections observed. As in most studies conducted on urinary tract infections, *Escherichia coli* were found to be the predominant isolate. This study did not find any significant difference in gender among children with urinary tract infection. The burden of urinary tract infections in young children living in developing countries remains undefined, despite being the most common bacterial infections in childhood. The diagnosis of urinary tract infection is important in young children since if left untreated can cause nephrourologic abnormalities and impaired renal function in future [37]. With an increase in the prevalence of...
The study had a number of limitations. Firstly, the study was conducted for the period of ten months and therefore could not assess seasonality of febrile episodes. Secondly, the study relied on verbal information provided by parents and guardians on the prior use of antimalarial drugs and antibiotics. This means that some of the children enrolled may well have been on antimicrobial therapy and this will have contributed to the low yield of positive cultures. In addition, the collection of only a single blood culture is known to reduce the possibility of isolating the organisms present in the bloodstream. The volume of blood inoculated into culture bottles was sometimes low due to difficulty in getting venous access especially from young patients and this might have contributed to the low bacterial yield. The study could not provide evidence-based data on respiratory and enteric viruses which are thought to cause majority of respiratory tract infections and diarrhoea in young children. Causative agents of zoonosis among outpatient children from this particular study community were not investigated. Understanding their prevalence would be of advantage despite not being investigated routinely at health facilities in the country.

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Author Contributions

Conceived and designed the experiments: CM BN JL AB PL ML ZL. Performed the experiments: CM AB PL. Analyzed the data: CM BN JL. Contributed reagents/materials/analysis tools: CM BN JL AB PL ML ZL. Contributed to the writing of the manuscript: CM. Reviewed the manuscript and provided critical inputs: CM BN JL AB PL ML ZL. Read and approved the final manuscript: CM BN JL AB PL ML ZL.

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